

Package ‘SummarizedExperiment’

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Title SummarizedExperiment container

Description The SummarizedExperiment container contains one or more assays, each represented by a matrix-like object of numeric or other mode. The rows typically represent genomic ranges of interest and the columns represent samples.

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Assays-class	<i>Assays objects</i>
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Description

The Assays virtual class and its methods provide a formal abstraction of the assays slot of [SummarizedExperiment](#) objects.

`SimpleListAssays` and `ShallowSimpleListAssays` are concrete subclasses of Assays with the latter being currently the default implementation of Assays objects. Other implementations (e.g. disk-based) could easily be added.

Note that these classes are not meant to be used directly by the end-user and the material in this man page is aimed at package developers.

Details

Assays objects have a list-like semantics with elements having matrix- or array-like semantics (e.g., `dim`, `dimnames`).

The Assays API consists of:

- (a) The `Assays()` constructor function.
- (b) Lossless back and forth coercion from/to [SimpleList](#). The coercion method from `SimpleList` doesn't need (and should not) validate the returned object.
- (c) `length`, `names`, ``names<-``, `[[`, ``[[<-``, `dim`, `[[<-``, `rbind`, `cbind`.

An Assays concrete subclass needs to implement (b) (required) plus, optionally any of the methods in (c).

IMPORTANT: Methods that return a modified Assays object (a.k.a. endomorphisms), that is, `[[` as well as replacement methods `names<-``, `[[<-``, and `[[<-``, must respect the *copy-on-change contract*. With objects that don't make use of references internally, the developer doesn't need to take any special action for that because it's automatically taken care of by R itself. However, for objects that do make use of references internally (e.g. environments, external pointers, pointer to a file on disk, etc...), the developer needs to be careful to implement endomorphisms with copy-on-change semantics. This can easily be achieved (and is what the default methods for Assays objects do) by performing a full (deep) copy of the object before modifying it instead of trying to modify it in-place. Note that the full (deep) copy is not always necessary in order to achieve copy-on-change semantics: it's enough (and often preferable for performance reasons) to copy only the parts of the objects that need to be modified.

Assays has currently 3 implementations which are formalized by concrete subclasses `SimpleListAssays`, `ShallowSimpleListAssays`, and `AssaysInEnv`. `ShallowSimpleListAssays` is the default.

`AssaysInEnv` is a *broken* alternative to `ShallowSimpleListAssays` that does NOT respect the *copy-on-change contract*. It is only provided for illustration purposes (see source file `Assays-class.R` for the details).

A little more detail about `ShallowSimpleListAssays`: a small reference class hierarchy (not exported from the **GenomicRanges** name space) defines a reference class `ShallowData` with a single field data of type ANY, and a derived class `ShallowSimpleListAssays` that specializes the type of data as `SimpleList`, and contains=c("ShallowData", "Assays"). The assays slot of a `SummarizedExperiment` object contains an instance of `ShallowSimpleListAssays`.

Author(s)

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See Also

- `SummarizedExperiment` objects.
- `SimpleList` objects in the `S4Vectors` package.

Examples

```
## -----
## DIRECT MANIPULATION OF Assays OBJECTS
## -----
m1 <- matrix(runif(24), ncol=3)
m2 <- matrix(runif(24), ncol=3)
a <- Assays(SimpleList(m1, m2))
a

as(a, "SimpleList")

length(a)
a[[2]]
dim(a)

b <- a[-4, 2]
b
length(b)
b[[2]]
dim(b)

names(a)
names(a) <- c("a1", "a2")
names(a)
a[["a2"]]

rbind(a, a)
cbind(a, a)

## -----
## COPY-ON-CHANGE CONTRACT
## -----

## ShallowSimpleListAssays objects have copy-on-change semantics but not
## AssaysInEnv objects. For example:
ssla <- as(SimpleList(m1, m2), "ShallowSimpleListAssays")
aie <- as(SimpleList(m1, m2), "AssaysInEnv")
```

```
## No names on 'ssla' and 'aie':
names(ssla)
names(aie)

ssla2 <- ssla
aie2 <- aie
names(ssla2) <- names(aie2) <- c("A1", "A2")

names(ssla) # still NULL (as expected)

names(aie) # changed! (because the names<-, AssaysInEnv method is not
# implemented in a way that respects the copy-on-change
# contract)
```

coverage-methods

Coverage of a RangedSummarizedExperiment object

Description

This man page documents the coverage method for [RangedSummarizedExperiment](#) objects.

Usage

```
## S4 method for signature 'RangedSummarizedExperiment'
coverage(x, shift=0L, width=NULL, weight=1L,
         method=c("auto", "sort", "hash"))
```

Arguments

`x` A [RangedSummarizedExperiment](#) object.
`shift`, `width`, `weight`, `method`
 See [?coverage](#) in the **GenomicRanges** package.

Details

This method operates on the `rowRanges` component of the [RangedSummarizedExperiment](#) object, which can be a [GenomicRanges](#) or [GRangesList](#) object.

More precisely, on [RangedSummarizedExperiment](#) object `x`, `coverage(x, ...)` is equivalent to `coverage(rowRanges(x), ...)`.

See [?coverage](#) in the **GenomicRanges** package for the details of how coverage operates on a [GenomicRanges](#) or [GRangesList](#) object.

Value

See [?coverage](#) in the **GenomicRanges** package.

See Also

- [RangedSummarizedExperiment](#) objects.
- The [coverage](#) man page in the **GenomicRanges** package where the coverage methods for [GenomicRanges](#) and [GRangesList](#) objects are documented.

Examples

```
nrows <- 20; ncols <- 6
counts <- matrix(runif(nrows * ncols, 1, 1e4), nrows)
rowRanges <- GRanges(rep(c("chr1", "chr2"), c(5, 15)),
                     IRanges(sample(1000L, 20), width=100),
                     strand=Rle(c("+", "-"), c(12, 8)),
                     seqlengths=c(chr1=1800, chr2=1300))
colData <- DataFrame(Treatment=rep(c("ChIP", "Input"), 3),
                    row.names=LETTERS[1:6])
rse <- SummarizedExperiment(assays=SimpleList(counts=counts),
                           rowRanges=rowRanges, colData=colData)

cvg <- coverage(rse)
cvg
stopifnot(identical(cvg, coverage(rowRanges(rse))))
```

findOverlaps-methods *Finding overlapping ranges in RangedSummarizedExperiment objects*

Description

This man page documents the `findOverlaps` methods for [RangedSummarizedExperiment](#) objects.

[RangedSummarizedExperiment](#) objects also support `countOverlaps`, `overlapsAny`, and `subsetByOverlaps` thanks to the default methods defined in the **IRanges** package and to the `findOverlaps` methods defined in this package and documented below.

Usage

```
## S4 method for signature 'RangedSummarizedExperiment,Vector'
findOverlaps(query, subject,
             maxgap=-1L, minoverlap=0L,
             type=c("any", "start", "end", "within", "equal"),
             select=c("all", "first", "last", "arbitrary"),
             ignore.strand=FALSE)
## S4 method for signature 'Vector,RangedSummarizedExperiment'
findOverlaps(query, subject,
             maxgap=-1L, minoverlap=0L,
             type=c("any", "start", "end", "within", "equal"),
             select=c("all", "first", "last", "arbitrary"),
             ignore.strand=FALSE)
```

Arguments

`query`, `subject` One of these two arguments must be a [RangedSummarizedExperiment](#) object.

`maxgap`, `minoverlap`, `type`

See [?findOverlaps](#) in the **GenomicRanges** package.

`select`, `ignore.strand`

See [?findOverlaps](#) in the **GenomicRanges** package.

Details

These methods operate on the `rowRanges` component of the `RangedSummarizedExperiment` object, which can be a `GenomicRanges` or `GRangesList` object.

More precisely, if any of the above functions is passed a `RangedSummarizedExperiment` object through the `query` and/or `subject` argument, then it behaves as if `rowRanges(query)` and/or `rowRanges(subject)` had been passed instead.

See `?findOverlaps` in the `GenomicRanges` package for the details of how `findOverlaps` and family operate on `GenomicRanges` and `GRangesList` objects.

Value

See `?findOverlaps` in the `GenomicRanges` package.

See Also

- `RangedSummarizedExperiment` objects.
- The `findOverlaps` man page in the `GenomicRanges` package where the `findOverlaps` family of methods for `GenomicRanges` and `GRangesList` objects is documented.

Examples

```
nrows <- 20; ncols <- 6
counts <- matrix(runif(nrows * ncols, 1, 1e4), nrows)
rowRanges <- GRanges(rep(c("chr1", "chr2"), c(5, 15)),
                     IRanges(sample(1000L, 20), width=100),
                     strand=Rle(c("+", "-"), c(12, 8)))
colData <- DataFrame(Treatment=rep(c("ChIP", "Input"), 3),
                    row.names=LETTERS[1:6])
rse0 <- SummarizedExperiment(assays=SimpleList(counts=counts),
                             rowRanges=rowRanges, colData=colData)
rse1 <- shift(rse0, 100)

hits <- findOverlaps(rse0, rse1)
hits
stopifnot(identical(hits, findOverlaps(rowRanges(rse0), rowRanges(rse1))))
stopifnot(identical(hits, findOverlaps(rse0, rowRanges(rse1))))
stopifnot(identical(hits, findOverlaps(rowRanges(rse0), rse1)))
```

inter-range-methods *Inter range transformations of a RangedSummarizedExperiment object*

Description

This man page documents the *inter range transformations* that are supported on `RangedSummarizedExperiment` objects.

Usage

```
## S4 method for signature 'RangedSummarizedExperiment'
isDisjoint(x, ignore.strand=FALSE)

## S4 method for signature 'RangedSummarizedExperiment'
disjointBins(x, ignore.strand=FALSE)
```

Arguments

`x` A [RangedSummarizedExperiment](#) object.

`ignore.strand` See [?isDisjoint](#) in the **GenomicRanges** package.

Details

These transformations operate on the `rowRanges` component of the [RangedSummarizedExperiment](#) object, which can be a [GenomicRanges](#) or [GRangesList](#) object.

More precisely, any of the above functions performs the following transformation on [RangedSummarizedExperiment](#) object `x`:

```
f(rowRanges(x), ...)
```

where `f` is the name of the function and `...` any additional arguments passed to it.

See [?isDisjoint](#) in the **GenomicRanges** package for the details of how these transformations operate on a [GenomicRanges](#) or [GRangesList](#) object.

Value

See [?isDisjoint](#) in the **GenomicRanges** package.

See Also

- [RangedSummarizedExperiment](#) objects.
- The [isDisjoint](#) man page in the **GenomicRanges** package where *inter range transformations* of a [GenomicRanges](#) or [GRangesList](#) object are documented.

Examples

```
nrows <- 20; ncols <- 6
counts <- matrix(runif(nrows * ncols, 1, 1e4), nrows)
rowRanges <- GRanges(rep(c("chr1", "chr2"), c(5, 15)),
                    IRanges(sample(1000L, 20), width=100),
                    strand=Rle(c("+", "-"), c(12, 8)))
colData <- DataFrame(Treatment=rep(c("ChIP", "Input"), 3),
                    row.names=LETTERS[1:6])
rse0 <- SummarizedExperiment(assays=SimpleList(counts=counts),
                             rowRanges=rowRanges, colData=colData)
rse1 <- shift(rse0, 99*start(rse0))

isDisjoint(rse0) # FALSE
isDisjoint(rse1) # TRUE

bins0 <- disjointBins(rse0)
bins0
```

```

stopifnot(identical(bins0, disjointBins(rowRanges(rse0))))

bins1 <- disjointBins(rse1)
bins1
stopifnot(all(bins1 == bins1[[1]]))

```

intra-range-methods *Intra range transformations of a RangedSummarizedExperiment object*

Description

This man page documents the *intra range transformations* that are supported on [RangedSummarizedExperiment](#) objects.

Usage

```

## S4 method for signature 'RangedSummarizedExperiment'
shift(x, shift=0L, use.names=TRUE)

## S4 method for signature 'RangedSummarizedExperiment'
narrow(x, start=NA, end=NA, width=NA, use.names=TRUE)

## S4 method for signature 'RangedSummarizedExperiment'
resize(x, width, fix="start", use.names=TRUE,
       ignore.strand=FALSE)

## S4 method for signature 'RangedSummarizedExperiment'
flank(x, width, start=TRUE, both=FALSE,
      use.names=TRUE, ignore.strand=FALSE)

## S4 method for signature 'RangedSummarizedExperiment'
promoters(x, upstream=2000, downstream=200)

## S4 method for signature 'RangedSummarizedExperiment'
restrict(x, start=NA, end=NA, keep.all.ranges=FALSE,
        use.names=TRUE)

## S4 method for signature 'RangedSummarizedExperiment'
trim(x, use.names=TRUE)

```

Arguments

`x` A [RangedSummarizedExperiment](#) object.

`shift`, `use.names` See [?shift](#) in the **GenomicRanges** package.

`start`, `end`, `width`, `fix` See [?shift](#) in the **GenomicRanges** package.

`ignore.strand`, `both` See [?shift](#) in the **GenomicRanges** package.

upstream, downstream

See [?shift](#) in the **GenomicRanges** package.

keep.all.ranges

See [?shift](#) in the **GenomicRanges** package.

Details

These transformations operate on the `rowRanges` component of the [RangedSummarizedExperiment](#) object, which can be a [GenomicRanges](#) or [GRangesList](#) object.

More precisely, any of the above functions performs the following transformation on [RangedSummarizedExperiment](#) object `x`:

```
rowRanges(x) <- f(rowRanges(x), ...)
```

where `f` is the name of the function and `...` any additional arguments passed to it.

See [?shift](#) in the **GenomicRanges** package for the details of how these transformations operate on a [GenomicRanges](#) or [GRangesList](#) object.

See Also

- [RangedSummarizedExperiment](#) objects.
- The [shift](#) man page in the **GenomicRanges** package where *intra range transformations* of a [GenomicRanges](#) or [GRangesList](#) object are documented.

Examples

```
nrows <- 20; ncols <- 6
counts <- matrix(runif(nrows * ncols, 1, 1e4), nrows)
rowRanges <- GRanges(rep(c("chr1", "chr2"), c(5, 15)),
                    IRanges(sample(1000L, 20), width=100),
                    strand=Rle(c("+", "-"), c(12, 8)))
colData <- DataFrame(Treatment=rep(c("ChIP", "Input"), 3),
                    row.names=LETTERS[1:6])
rse0 <- SummarizedExperiment(assays=SimpleList(counts=counts),
                            rowRanges=rowRanges, colData=colData)
```

```
rse1 <- shift(rse0, 1)
stopifnot(identical(
  rowRanges(rse1),
  shift(rowRanges(rse0), 1)
))
```

```
se2 <- narrow(rse0, start=10, end=-15)
stopifnot(identical(
  rowRanges(se2),
  narrow(rowRanges(rse0), start=10, end=-15)
))
```

```
se3 <- resize(rse0, width=75)
stopifnot(identical(
  rowRanges(se3),
  resize(rowRanges(rse0), width=75)
))
```

```
se4 <- flank(rse0, width=20)
```

```

stopifnot(identical(
  rowRanges(se4),
  flank(rowRanges(rse0), width=20)
))

se5 <- restrict(rse0, start=200, end=700, keep.all.ranges=TRUE)
stopifnot(identical(
  rowRanges(se5),
  restrict(rowRanges(rse0), start=200, end=700, keep.all.ranges=TRUE)
))

```

```
makeSummarizedExperimentFromDataFrame
```

Make a RangedSummarizedExperiment from a data.frame or DataFrame

Description

`makeSummarizedExperimentFromDataFrame` uses `data.frame` or `DataFrame` column names to create a `GRanges` object for the `rowRanges` of the resulting `SummarizedExperiment` object. It requires that non-range data columns be coercible into a numeric matrix for the `SummarizedExperiment` constructor. All columns that are not part of the row ranges attribute are assumed to be experiment data; thus, keeping metadata columns will not be supported. Note that this function only returns `SummarizedExperiment` objects with a single assay.

If metadata columns are to be kept, one can first construct the row ranges attribute by using the `makeGRangesFromDataFrame` function and subsequently creating the `SummarizedExperiment`.

Usage

```

makeSummarizedExperimentFromDataFrame(df,
                                     ...,
                                     seqinfo = NULL,
                                     starts.in.df.are.0based = FALSE)

```

Arguments

<code>df</code>	A <code>data.frame</code> or <code>DataFrame</code> object. If not, then the function first tries to turn <code>df</code> into a data frame with <code>as.data.frame(df)</code> .
<code>...</code>	Additional arguments passed on to <code>makeGRangesFromDataFrame</code>
<code>seqinfo</code>	Either <code>NULL</code> , or a <code>Seqinfo</code> object, or a character vector of seqlevels, or a named numeric vector of sequence lengths. When not <code>NULL</code> , it must be compatible with the genomic ranges in <code>df</code> i.e. it must include at least the sequence levels represented in <code>df</code> .
<code>starts.in.df.are.0based</code>	<code>TRUE</code> or <code>FALSE</code> (the default). If <code>TRUE</code> , then the start positions of the genomic ranges in <code>df</code> are considered to be <i>0-based</i> and are converted to <i>1-based</i> in the returned <code>GRanges</code> object. This feature is intended to make it more convenient to handle input that contains data obtained from resources using the "0-based start" convention. A notorious example of such resource is the UCSC Table Browser (http://genome.ucsc.edu/cgi-bin/hgTables).

Value

A [RangedSummarizedExperiment](#) object with rowRanges and a single assay

Author(s)

M. Ramos

See Also

- [makeGRangesFromDataFrame](#)

Examples

```
## -----
## BASIC EXAMPLES
## -----

# Note that rownames of the data.frame are also rownames of the result
df <- data.frame(chr="chr2", start = 11:15, end = 12:16,
  strand = c("+", "-", "+", "*", "."), expr0 = 3:7,
  expr1 = 8:12, expr2 = 12:16,
  row.names = paste0("GENE", letters[5:1]))

df

exRSE <- makeSummarizedExperimentFromDataFrame(df)

exRSE

assay(exRSE)

rowRanges(exRSE)
```

```
makeSummarizedExperimentFromExpressionSet
```

Make a RangedSummarizedExperiment object from an ExpressionSet and vice-versa

Description

Coercion between [RangedSummarizedExperiment](#) and [ExpressionSet](#) is supported in both directions.

For going from [ExpressionSet](#) to [RangedSummarizedExperiment](#), the `makeSummarizedExperimentFromExpressionSet` function is also provided to let the user control how to map features to ranges.

Usage

```
makeSummarizedExperimentFromExpressionSet(from,
  mapFun=naiveRangeMapper,
  ...)

## range mapping functions
naiveRangeMapper(from)
probeRangeMapper(from)
geneRangeMapper(txDbPackage, key = "ENTREZID")
```

Arguments

from	An ExpressionSet object.
mapFun	A function which takes an ExpressionSet object and returns a GRanges , or GRangesList object which corresponds to the genomic ranges used in the ExpressionSet . The rownames of the returned GRanges are used to match the featureNames of the ExpressionSet . The naiveRangeMapper function is used by default.
...	Additional arguments passed to mapFun .
txDbPackage	A character string with the Transcript Database to use for the mapping.
key	A character string with the Gene key to use for the mapping.

Value

`makeSummarizedExperimentFromExpressionSet` takes an [ExpressionSet](#) object as input and a *range mapping function* that maps the features to ranges. It then returns a [RangedSummarizedExperiment](#) object that corresponds to the input.

The range mapping functions return a [GRanges](#) object, with the [rownames](#) corresponding to the [featureNames](#) of the [ExpressionSet](#) object.

Author(s)

Jim Hester, james.f.hester@gmail.com

See Also

- [RangedSummarizedExperiment](#) objects.
- [ExpressionSet](#) objects in the **Biobase** package.
- [TxDb](#) objects in the **GenomicFeatures** package.

Examples

```
## -----
## GOING FROM ExpressionSet TO SummarizedExperiment
## -----

data(sample.ExpressionSet, package="Biobase")

# naive coercion
makeSummarizedExperimentFromExpressionSet(sample.ExpressionSet)
as(sample.ExpressionSet, "RangedSummarizedExperiment")
as(sample.ExpressionSet, "SummarizedExperiment")

# using probe range mapper
makeSummarizedExperimentFromExpressionSet(sample.ExpressionSet, probeRangeMapper)

# using the gene range mapper
makeSummarizedExperimentFromExpressionSet(sample.ExpressionSet,
                                           geneRangeMapper("TxDb.Hsapiens.UCSC.hg19.knownGene"))

## -----
## GOING FROM SummarizedExperiment TO ExpressionSet
## -----
```

```
example(RangedSummarizedExperiment) # to create 'rse'
rse
as(rse, "ExpressionSet")
```

nearest-methods	<i>Finding the nearest range neighbor in RangedSummarizedExperiment objects</i>
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Description

This man page documents the nearest methods and family (i.e. precede, follow, distance, and distanceToNearest methods) for [RangedSummarizedExperiment](#) objects.

Usage

```
## S4 method for signature 'RangedSummarizedExperiment,ANY'
precede(x, subject, select=c("arbitrary", "all"),
        ignore.strand=FALSE)
## S4 method for signature 'ANY,RangedSummarizedExperiment'
precede(x, subject, select=c("arbitrary", "all"),
        ignore.strand=FALSE)

## S4 method for signature 'RangedSummarizedExperiment,ANY'
follow(x, subject, select=c("arbitrary", "all"),
        ignore.strand=FALSE)
## S4 method for signature 'ANY,RangedSummarizedExperiment'
follow(x, subject, select=c("arbitrary", "all"),
        ignore.strand=FALSE)

## S4 method for signature 'RangedSummarizedExperiment,ANY'
nearest(x, subject, select=c("arbitrary", "all"), ignore.strand=FALSE)
## S4 method for signature 'ANY,RangedSummarizedExperiment'
nearest(x, subject, select=c("arbitrary", "all"), ignore.strand=FALSE)

## S4 method for signature 'RangedSummarizedExperiment,ANY'
distance(x, y, ignore.strand=FALSE, ...)
## S4 method for signature 'ANY,RangedSummarizedExperiment'
distance(x, y, ignore.strand=FALSE, ...)

## S4 method for signature 'RangedSummarizedExperiment,ANY'
distanceToNearest(x, subject, ignore.strand=FALSE, ...)
## S4 method for signature 'ANY,RangedSummarizedExperiment'
distanceToNearest(x, subject, ignore.strand=FALSE, ...)
```

Arguments

x, subject One of these two arguments must be a [RangedSummarizedExperiment](#) object.
select, ignore.strand
See [?nearest](#) in the **GenomicRanges** package.

y For the distance methods, one of x or y must be a [RangedSummarizedExperiment](#) object.

... Additional arguments for methods.

Details

These methods operate on the `rowRanges` component of the [RangedSummarizedExperiment](#) object, which can be a [GenomicRanges](#) or [GRangesList](#) object.

More precisely, if any of the above functions is passed a [RangedSummarizedExperiment](#) object through the `x`, `subject`, and/or `y` argument, then it behaves as if `rowRanges(x)`, `rowRanges(subject)`, and/or `rowRanges(y)` had been passed instead.

See [?nearest](#) in the **GenomicRanges** package for the details of how `nearest` and family operate on [GenomicRanges](#) and [GRangesList](#) objects.

Value

See [?nearest](#) in the **GenomicRanges** package.

See Also

- [RangedSummarizedExperiment](#) objects.
- The [nearest](#) man page in the **GenomicRanges** package where the nearest family of methods for [GenomicRanges](#) and [GRangesList](#) objects is documented.

Examples

```
nrows <- 20; ncols <- 6
counts <- matrix(runif(nrows * ncols, 1, 1e4), nrows)
rowRanges <- GRanges(rep(c("chr1", "chr2"), c(5, 15)),
                    IRanges(sample(1000L, 20), width=100),
                    strand=Rle(c("+", "-"), c(12, 8)))
colData <- DataFrame(Treatment=rep(c("ChIP", "Input"), 3),
                    row.names=LETTERS[1:6])
rse0 <- SummarizedExperiment(assays=SimpleList(counts=counts),
                             rowRanges=rowRanges, colData=colData)
rse1 <- shift(rse0, 100)

res <- nearest(rse0, rse1)
res
stopifnot(identical(res, nearest(rowRanges(rse0), rowRanges(rse1))))
stopifnot(identical(res, nearest(rse0, rowRanges(rse1))))
stopifnot(identical(res, nearest(rowRanges(rse0), rse1)))

res <- nearest(rse0) # missing subject
res
stopifnot(identical(res, nearest(rowRanges(rse0))))

hits <- nearest(rse0, rse1, select="all")
hits
stopifnot(identical(
  hits,
  nearest(rowRanges(rse0), rowRanges(rse1), select="all")
))
stopifnot(identical(
  hits,
```

```

    nearest(rse0, rowRanges(rse1), select="all")
  ))
  stopifnot(identical(
    hits,
    nearest(rowRanges(rse0), rse1, select="all")
  ))

```

RangedSummarizedExperiment-class

RangedSummarizedExperiment objects

Description

The `RangedSummarizedExperiment` class is a matrix-like container where rows represent ranges of interest (as a `GRanges` or `GRangesList` object) and columns represent samples (with sample data summarized as a `DataFrame`). A `RangedSummarizedExperiment` contains one or more assays, each represented by a matrix-like object of numeric or other mode.

`RangedSummarizedExperiment` is a subclass of `SummarizedExperiment` and, as such, all the methods documented in `?SummarizedExperiment` also work on a `RangedSummarizedExperiment` object. The methods documented below are additional methods that are specific to `RangedSummarizedExperiment` objects.

Usage

```

## Constructor

SummarizedExperiment(assays, ...)
## S4 method for signature 'SimpleList'
SummarizedExperiment(assays, rowData=NULL, rowRanges=GRangesList(),
  colData=DataFrame(), metadata=list())
## S4 method for signature 'ANY'
SummarizedExperiment(assays, ...)
## S4 method for signature 'list'
SummarizedExperiment(assays, ...)
## S4 method for signature 'missing'
SummarizedExperiment(assays, ...)

## Accessors

rowRanges(x, ...)
rowRanges(x, ...) <- value

## Subsetting

## S4 method for signature 'RangedSummarizedExperiment'
subset(x, subset, select, ...)

## rowRanges access
## see 'GRanges compatibility', below

```

Arguments

assays	A list or SimpleList of matrix-like elements, or a matrix-like object. All elements of the list must have the same dimensions, and dimension names (if present) must be consistent across elements and with the row names of rowRanges and colData.
rowData	A DataFrame object describing the rows. Row names, if present, become the row names of the SummarizedExperiment object. The number of rows of the DataFrame must equal the number of rows of the matrices in assays.
rowRanges	A GRanges or GRangesList object describing the ranges of interest. Names, if present, become the row names of the SummarizedExperiment object. The length of the GRanges or GRangesList must equal the number of rows of the matrices in assays. If rowRanges is missing, a SummarizedExperiment instance is returned.
colData	An optional DataFrame describing the samples. Row names, if present, become the column names of the RangedSummarizedExperiment.
metadata	An optional list of arbitrary content describing the overall experiment.
...	For SummarizedExperiment, S4 methods list and matrix, arguments identical to those of the SimpleList method. For rowRanges, ignored.
x	A RangedSummarizedExperiment object. The rowRanges setter will also accept a SummarizedExperiment object and will first coerce it to RangedSummarizedExperiment before it sets value on it.
value	A GRanges or GRangesList object.
subset	An expression which, when evaluated in the context of rowRanges(x), is a logical vector indicating elements or rows to keep: missing values are taken as false.
select	An expression which, when evaluated in the context of colData(x), is a logical vector indicating elements or rows to keep: missing values are taken as false.

Details

The rows of a RangedSummarizedExperiment object represent ranges (in genomic coordinates) of interest. The ranges of interest are described by a GRanges or a GRangesList object, accessible using the rowRanges function, described below. The GRanges and GRangesList classes contains sequence (e.g., chromosome) name, genomic coordinates, and strand information. Each range can be annotated with additional data; this data might be used to describe the range or to summarize results (e.g., statistics of differential abundance) relevant to the range. Rows may or may not have row names; they often will not.

Constructor

RangedSummarizedExperiment instances are constructed using the SummarizedExperiment function with arguments outlined above.

Accessors

In the following code snippets, x is a RangedSummarizedExperiment object.

```
rowRanges(x), rowRanges(x) <- value: Get or set the row data. value is a GenomicRanges object. Row names of value must be NULL or consistent with the existing row names of x.
```


GRanges compatibility (rowRanges access)

Many [GRanges](#) and [GRangesList](#) operations are supported on `RangedSummarizedExperiment` objects, using `rowRanges`.

Supported operations include: `pcompare`, `duplicated`, `end`, `end<-`, `granges`, `is.unsorted`, `match`, `mcols`, `mcols<-`, `order`, `ranges`, `ranges<-`, `rank`, `seqinfo`, `seqinfo<-`, `seqnames`, `sort`, `start`, `start<-`, `strand`, `strand<-`, `width`, `width<-`.

See also `?shift`, `?isDisjoint`, `?coverage`, `?findOverlaps`, and `?nearest` for more *GRanges compatibility methods*.

Not all [GRanges](#) operations are supported, because they do not make sense for `RangedSummarizedExperiment` objects (e.g., `length`, `name`, `as.data.frame`, `c`, `splitAsList`), involve non-trivial combination or splitting of rows (e.g., `disjoin`, `gaps`, `reduce`, `unique`), or have not yet been implemented (`Ops`, `map`, `window`, `window<-`).

Subsetting

In the code snippets below, `x` is a `RangedSummarizedExperiment` object.

`subset(x, subset, select)`: Create a subset of `x` using an expression `subset` referring to columns of `rowRanges(x)` (including `'seqnames'`, `'start'`, `'end'`, `'width'`, `'strand'`, and `names(rowData(x))`) and / or `select` referring to column names of `colData(x)`.

Extension

`RangedSummarizedExperiment` is implemented as an S4 class, and can be extended in the usual way, using `contains="RangedSummarizedExperiment"` in the new class definition.

Author(s)

Martin Morgan, mtmorgan@fhcrc.org

See Also

- [SummarizedExperiment](#) objects.
- `shift`, `isDisjoint`, `coverage`, `findOverlaps`, and `nearest` for more *GRanges compatibility methods*.
- [GRanges](#) objects in the **GenomicRanges** package.

Examples

```
nrows <- 200; ncols <- 6
counts <- matrix(runif(nrows * ncols, 1, 1e4), nrows)
rowRanges <- GRanges(rep(c("chr1", "chr2"), c(50, 150)),
                    IRanges(floor(runif(200, 1e5, 1e6)), width=100),
                    strand=sample(c("+", "-"), 200, TRUE),
                    feature_id=sprintf("ID%03d", 1:200))
colData <- DataFrame(Treatment=rep(c("ChIP", "Input"), 3),
                    row.names=LETTERS[1:6])
rse <- SummarizedExperiment(assays=SimpleList(counts=counts),
                          rowRanges=rowRanges, colData=colData)
rse
dim(rse)
dimnames(rse)
assayNames(rse)
```

```

head(assay(rse))
assays(rse) <- endoapply(assays(rse), asinh)
head(assay(rse))

rowRanges(rse)
rowData(rse) # same as 'mcols(rowRanges(rse))'
colData(rse)

rse[, rse$Treatment == "ChIP"]

## cbind() combines objects with the same ranges but different samples:
rse1 <- rse
rse2 <- rse1[,1:3]
colnames(rse2) <- letters[1:ncol(rse2)]
cmb1 <- cbind(rse1, rse2)
dim(cmb1)
dimnames(cmb1)

## rbind() combines objects with the same samples but different ranges:
rse1 <- rse
rse2 <- rse1[1:50,]
rownames(rse2) <- letters[1:nrow(rse2)]
cmb2 <- rbind(rse1, rse2)
dim(cmb2)
dimnames(cmb2)

## Coercion to/from SummarizedExperiment:
se0 <- as(rse, "SummarizedExperiment")
se0

as(se0, "RangedSummarizedExperiment")

## Setting rowRanges on a SummarizedExperiment object turns it into a
## RangedSummarizedExperiment object:
se <- se0
rowRanges(se) <- rowRanges
se # RangedSummarizedExperiment

## Sanity checks:
stopifnot(identical(assays(se0), assays(rse)))
stopifnot(identical(dim(se0), dim(rse)))
stopifnot(identical(dimnames(se0), dimnames(rse)))
stopifnot(identical(rowData(se0), rowData(rse)))
stopifnot(identical(colData(se0), colData(rse)))

```

readKallisto

Input kallisto or kallisto bootstrap results.

Description

readKallisto inputs several kallisto output files into a single SummarizedExperiment instance, with rows corresponding to estimated transcript abundance and columns to samples. readKallistoBootstrap inputs kallisto bootstrap replicates of a single sample into a matrix of transcript x bootstrap abundance estimates.

Usage

```
readKallisto(files,
             json = file.path(dirname(files), "run_info.json"),
             h5 = any(grepl("\\.h5$", files)), what = KALLISTO_ASSAYS,
             as = c("SummarizedExperiment", "list", "matrix"))

readKallistoBootstrap(file, i, j)
```

Arguments

files	character() paths to kallisto ‘abundance.tsv’ output files. The assumption is that files are organized in the way implied by kallisto, with each sample in a distinct directory, and the directory containing files abundance.tsv, run_info.json, and perhaps abundance.h5.
json	character() vector of the same length as files specifying the location of JSON files produced by kallisto and containing information on the run. The default assumes that json files are in the same directory as the corresponding abundance file.
h5	character() vector of the same length as files specifying the location of HDF5 files produced by kallisto and containing bootstrap estimates. The default assumes that HDF5 files are in the same directory as the corresponding abundance file.
what	character() vector of kallisto per-sample outputs to be input. See KALLISTO_ASSAYS for available values.
as	character(1) specifying the output format. See Value for additional detail.
file	character(1) path to a single HDF5 output file.
i, j	integer() vector of row (i) and column (j) indexes to input.

Value

A SummarizedExperiment, list, or matrix, depending on the value of argument as; by default a SummarizedExperiment. The as="SummarizedExperiment" rowData(se) the length of each transcript; colData(se) includes summary information on each sample, including the number of targets and bootstraps, the kallisto and index version, the start time and operating system call used to create the file. assays() contains one or more transcript x sample matrices of parameters estimated by kallisto (see KALLISTO_ASSAYS).

as="list" return value contains information similar to SummarizedExperiment with row, column and assay data as elements of the list without coordination of row and column annotations into an integrated data container. as="matrix" returns the specified assay as a simple R matrix.

Author(s)

Martin Morgan martin.morgan@roswellpark.org

References

<http://pachterlab.github.io/kallisto> software for quantifying transcript abundance.

Examples

```

outputs <- system.file(package="SummarizedExperiment", "extdata",
  "kallisto")
files <- dir(outputs, pattern="abundance.tsv", full=TRUE, recursive=TRUE)
stopifnot(all(file.exists(files)))

## default: input 'est_counts'
(se <- readKallisto(files, as="SummarizedExperiment"))
str(readKallisto(files, as="list"))
str(readKallisto(files, as="matrix"))

## available assays
KALLISTO_ASSAYS
## one or more assay
readKallisto(files, what=c("tpm", "eff_length"))

## alternatively: read hdf5 files
files <- sub(".tsv", ".h5", files, fixed=TRUE)
readKallisto(files)

## input all bootstraps
xx <- readKallistoBootstrap(files[1])
ridx <- head(which(rowSums(xx) != 0), 3)
cidx <- c(1:5, 96:100)
xx[ridx, cidx]

## selective input of rows (transcripts) and/or bootstraps
readKallistoBootstrap(files[1], i=c(ridx, rev(ridx)), j=cidx)

```

```
saveHDF5SummarizedExperiment
```

Save/load a HDF5-based SummarizedExperiment object

Description

saveHDF5SummarizedExperiment and loadHDF5SummarizedExperiment can be used to save/load a HDF5-based [SummarizedExperiment](#) object to/from disk.

Usage

```

saveHDF5SummarizedExperiment(x, dir="my_h5_se", replace=FALSE,
  chunk_dim=NULL, level=NULL, verbose=FALSE)
loadHDF5SummarizedExperiment(dir="my_h5_se")

```

Arguments

x	A SummarizedExperiment object.
dir	The path (as a single string) to the directory where to save the HDF5-based SummarizedExperiment object or to load it from. When saving, the directory will be created so should not already exist, unless replace is set to TRUE.
replace	If directory dir already exists, should it be replaced with a new one? The content of the existing directory will be lost!

chunk_dim, level	The dimensions of the chunks and the compression level to use for writing the assay data to disk. Passed to the internal calls to <code>HDF5Array::writeHDF5Array</code> . See <code>?HDF5Array::writeHDF5Array</code> for more information.
verbose	Set to TRUE to make the function display progress.

Details

These functions use functionalities from the **rhdf5** and **HDF5Array** packages internally and so require these packages to be installed.

`saveHDF5SummarizedExperiment` creates the directory specified thru the `dir` argument and then populates it with the HDF5 datasets (one per assay in `x`) plus a serialized version of `x` that contains pointers to these datasets. This directory provides a self-contained HDF5-based representation of `x` that can then be loaded back in R with `loadHDF5SummarizedExperiment`. Note that this directory is *relocatable* i.e. it can be moved (or copied) to a different place, on the same or a different computer, before calling `loadHDF5SummarizedExperiment` on it. For convenient sharing with collaborators, it is suggested to turn it into a tarball (with Unix command `tar`), or zip file, before the transfer. Please keep in mind that `saveHDF5SummarizedExperiment` and `loadHDF5SummarizedExperiment` don't know how to produce/read tarballs or zip files at the moment, so the process of packaging/extracting the tarball or zip file is entirely the user responsibility. It is typically done from outside R.

Finally please note that, depending on the size of the data to write to disk and the performance of the disk, `saveHDF5SummarizedExperiment` can take a long time to complete. Use `verbose=TRUE` to see its progress.

`loadHDF5SummarizedExperiment` is generally very fast, even if the assay data is big, because all the assays in the returned object are **HDF5Array** objects pointing to the on-disk HDF5 datasets located in `dir`. **HDF5Array** objects are typically light-weight in memory.

Value

`saveHDF5SummarizedExperiment` returns an invisible **SummarizedExperiment** object where all the assays are **HDF5Array** objects pointing to the HDF5 datasets saved in `dir`. It's in fact the same object as the object that would be returned by calling `loadHDF5SummarizedExperiment` on `dir`.

Author(s)

Hervé Pagès

See Also

- **SummarizedExperiment** and **RangedSummarizedExperiment** objects.
- **HDF5Array** objects in the **HDF5Array** package.
- The `writeHDF5Array` function in the **HDF5Array** package, which `saveHDF5SummarizedExperiment` uses internally to write the assay data to disk.

Examples

```
nrows <- 200; ncols <- 6
counts <- matrix(runif(nrows * ncols, 1, 1e4), nrows)
colData <- DataFrame(Treatment=rep(c("ChIP", "Input"), 3),
  row.names=LETTERS[1:6])
se0 <- SummarizedExperiment(assays=SimpleList(counts=counts),
```

```

                                colData=colData)
se0

## Save 'se0' as an HDF5-based SummarizedExperiment object:
dir <- sub("file", "h5_se0_", tempfile())
h5_se0 <- saveHDF5SummarizedExperiment(se0, dir)
h5_se0
assay(h5_se0, withDimnames=FALSE) # HDF5Matrix object

h5_se0b <- loadHDF5SummarizedExperiment(dir)
h5_se0b
assay(h5_se0b, withDimnames=FALSE) # HDF5Matrix object

## Sanity checks:
stopifnot(is(assay(h5_se0, withDimnames=FALSE), "HDF5Matrix"))
stopifnot(all(DelayedArray(assay(se0)) == assay(h5_se0)))
stopifnot(is(assay(h5_se0b, withDimnames=FALSE), "HDF5Matrix"))
stopifnot(all(DelayedArray(assay(se0)) == assay(h5_se0b)))

## -----
## More sanity checks
## -----

## Make a copy of directory 'dir':
somedir <- sub("file", "somedir", tempfile())
dir.create(somedir)
file.copy(dir, somedir, recursive=TRUE)
dir2 <- list.files(somedir, full.names=TRUE)

## 'dir2' contains a copy of 'dir'. Call loadHDF5SummarizedExperiment()
## on it.
h5_se0c <- loadHDF5SummarizedExperiment(dir2)

stopifnot(is(assay(h5_se0c, withDimnames=FALSE), "HDF5Matrix"))
stopifnot(all(DelayedArray(assay(se0)) == assay(h5_se0c)))

```

SummarizedExperiment-class

SummarizedExperiment objects

Description

The SummarizedExperiment class is a matrix-like container where rows represent features of interest (e.g. genes, transcripts, exons, etc...) and columns represent samples (with sample data summarized as a [DataFrame](#)). A SummarizedExperiment object contains one or more assays, each represented by a matrix-like object of numeric or other mode.

Note that SummarizedExperiment is the parent of the [RangedSummarizedExperiment](#) class which means that all the methods documented below also work on a [RangedSummarizedExperiment](#) object.

Usage

```

## Constructor

# See ?RangedSummarizedExperiment for the constructor function.

## Accessors

assayNames(x, ...)
assayNames(x, ...) <- value
assays(x, ..., withDimnames=TRUE)
assays(x, ..., withDimnames=TRUE) <- value
assay(x, i, ...)
assay(x, i, ...) <- value
rowData(x, ...)
rowData(x, ...) <- value
colData(x, ...)
colData(x, ...) <- value
#dim(x)
#dimnames(x)
#dimnames(x) <- value

## Quick colData access

## S4 method for signature 'SummarizedExperiment'
x$name
## S4 replacement method for signature 'SummarizedExperiment'
x$name <- value
## S4 method for signature 'SummarizedExperiment,ANY,missing'
x[[i, j, ...]]
## S4 replacement method for signature 'SummarizedExperiment,ANY,missing'
x[[i, j, ...]] <- value

## Subsetting

## S4 method for signature 'SummarizedExperiment'
x[i, j, ..., drop=TRUE]
## S4 replacement method for signature 'SummarizedExperiment,ANY,ANY,SummarizedExperiment'
x[i, j] <- value

## Combining

## S4 method for signature 'SummarizedExperiment'
cbind(..., deparse.level=1)
## S4 method for signature 'SummarizedExperiment'
rbind(..., deparse.level=1)

## On-disk realization
## S4 method for signature 'SummarizedExperiment'
realize(x)

```

Arguments

x A SummarizedExperiment object.

...	For assay, ... may contain withDimnames, which is forwarded to assays. For rowData, arguments passed thru ... are forwarded to <code>mcols</code> . For cbind, rbind, ... contains SummarizedExperiment objects to be combined. For other accessors, ignored.
i, j	For assay, assay<-, i is an integer or numeric scalar; see 'Details' for additional constraints. For [, SummarizedExperiment, [, SummarizedExperiment<-, i, j are subscripts that can act to subset the rows and columns of x, that is the matrix elements of assays. For [[, SummarizedExperiment, [[<-, SummarizedExperiment, i is a scalar index (e.g., character(1) or integer(1)) into a column of colData.
name	A symbol representing the name of a column of colData.
withDimnames	A logical(1), indicating whether dimnames should be applied to extracted assay elements. Setting withDimnames=FALSE increases the speed and memory efficiency with which assays are extracted. withDimnames=TRUE in the getter assays<- allows efficient complex assignments (e.g., updating names of assays, names(assays(x, withDimnames=FALSE)) = ... is more efficient than names(assays(x)) = ...); it does not influence actual assignment of dimnames to assays.
drop	A logical(1), ignored by these methods.
value	An object of a class specified in the S4 method signature or as outlined in 'Details'.
deparse.level	See ?base::cbind for a description of this argument.

Details

The SummarizedExperiment class is meant for numeric and other data types derived from a sequencing experiment. The structure is rectangular like a matrix, but with additional annotations on the rows and columns, and with the possibility to manage several assays simultaneously.

The rows of a SummarizedExperiment object represent features of interest. Information about these features is stored in a [DataFrame](#) object, accessible using the function `rowData`. The [DataFrame](#) must have as many rows as there are rows in the SummarizedExperiment object, with each row of the [DataFrame](#) providing information on the feature in the corresponding row of the SummarizedExperiment object. Columns of the [DataFrame](#) represent different attributes of the features of interest, e.g., gene or transcript IDs, etc.

Each column of a SummarizedExperiment object represents a sample. Information about the samples are stored in a [DataFrame](#), accessible using the function `colData`, described below. The [DataFrame](#) must have as many rows as there are columns in the SummarizedExperiment object, with each row of the [DataFrame](#) providing information on the sample in the corresponding column of the SummarizedExperiment object. Columns of the [DataFrame](#) represent different sample attributes, e.g., tissue of origin, etc. Columns of the [DataFrame](#) can themselves be annotated (via the `mcols` function). Column names typically provide a short identifier unique to each sample.

A SummarizedExperiment object can also contain information about the overall experiment, for instance the lab in which it was conducted, the publications with which it is associated, etc. This information is stored as a list object, accessible using the `metadata` function. The form of the data associated with the experiment is left to the discretion of the user.

The SummarizedExperiment container is appropriate for matrix-like data. The data are accessed using the `assays` function, described below. This returns a [SimpleList](#) object. Each element of

the list must itself be a matrix (of any mode) and must have dimensions that are the same as the dimensions of the SummarizedExperiment in which they are stored. Row and column names of each matrix must either be NULL or match those of the SummarizedExperiment during construction. It is convenient for the elements of [SimpleList](#) of assays to be named.

Constructor

SummarizedExperiment instances are constructed using the SummarizedExperiment function documented in [?RangedSummarizedExperiment](#).

Accessors

In the following code snippets, `x` is a SummarizedExperiment object.

`assays(x), assays(x) <- value`: Get or set the assays. `value` is a list or SimpleList, each element of which is a matrix with the same dimensions as `x`.

`assay(x, i), assay(x, i) <- value`: A convenient alternative (to `assays(x)[[i]]`, `assays(x)[[i]] <- value`) to get or set the `i`th (default first) assay element. `value` must be a matrix of the same dimension as `x`, and with dimension names NULL or consistent with those of `x`.

`assayNames(x), assayNames(x) <- value`: Get or set the names of `assay()` elements.

`rowData(x), rowData(x) <- value`: Get or set the row data. `value` is a [DataFrame](#) object. Row names of `value` must be NULL or consistent with the existing row names of `x`.

`colData(x), colData(x) <- value`: Get or set the column data. `value` is a [DataFrame](#) object. Row names of `value` must be NULL or consistent with the existing column names of `x`.

`metadata(x), metadata(x) <- value`: Get or set the experiment data. `value` is a list with arbitrary content.

`dim(x)`: Get the dimensions (features of interest x samples) of the SummarizedExperiment.

`dimnames(x), dimnames(x) <- value`: Get or set the dimension names. `value` is usually a list of length 2, containing elements that are either NULL or vectors of appropriate length for the corresponding dimension. `value` can be NULL, which removes dimension names. This method implies that `rownames`, `rownames<-`, `colnames`, and `colnames<-` are all available.

Subsetting

In the code snippets below, `x` is a SummarizedExperiment object.

`x[i, j], x[i, j] <- value`: Create or replace a subset of `x`. `i, j` can be numeric, logical, character, or missing. `value` must be a SummarizedExperiment object with dimensions, dimension names, and assay elements consistent with the subset `x[i, j]` being replaced.

Additional subsetting accessors provide convenient access to `colData` columns

`x$name, x$name <- value` Access or replace column name in `x`.

`x[[i, ...]], x[[i, ...]] <- value` Access or replace column `i` in `x`.

Combining

In the code snippets below, `...` are SummarizedExperiment objects to be combined.

`cbind(...)`: `cbind` combines objects with the same features of interest but different samples (columns in assays). The `colnames` in `colData(SummarizedExperiment)` must match or an error is thrown. Duplicate columns of `rowData(SummarizedExperiment)` must contain the same data.

Data in assays are combined by name matching; if all assay names are `NULL` matching is by position. A mixture of names and `NULL` throws an error.

metadata from all objects are combined into a list with no name checking.

`rbind(...)`: `rbind` combines objects with the same samples but different features of interest (rows in assays). The `colnames` in `rowData(SummarizedExperiment)` must match or an error is thrown. Duplicate columns of `colData(SummarizedExperiment)` must contain the same data.

Data in assays are combined by name matching; if all assay names are `NULL` matching is by position. A mixture of names and `NULL` throws an error.

metadata from all objects are combined into a list with no name checking.

Implementation and Extension

This section contains advanced material meant for package developers.

`SummarizedExperiment` is implemented as an S4 class, and can be extended in the usual way, using `contains="SummarizedExperiment"` in the new class definition.

In addition, the representation of the assays slot of `SummarizedExperiment` is as a virtual class `Assays`. This allows derived classes (`contains="Assays"`) to easily implement alternative requirements for the assays, e.g., backed by file-based storage like `NetCDF` or the `ff` package, while re-using the existing `SummarizedExperiment` class without modification. See [Assays](#) for more information.

The current assays slot is implemented as a reference class that has copy-on-change semantics. This means that modifying non-assay slots does not copy the (large) assay data, and at the same time the user is not surprised by reference-based semantics. Updates to non-assay slots are very fast; updating the assays slot itself can be 5x or more faster than with an S4 instance in the slot. One useful technique when working with assay or assays function is use of the `withDimnames=FALSE` argument, which benefits speed and memory use by not copying dimnames from the row- and colData elements to each assay.

Author(s)

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See Also

- [RangedSummarizedExperiment](#) objects.
- [DataFrame](#), [SimpleList](#), and [Annotated](#) objects in the `S4Vectors` package.
- The `metadata` and `mcols` accessors in the `S4Vectors` package.
- [saveHDF5SummarizedExperiment](#) and [loadHDF5SummarizedExperiment](#) for saving/loading a HDF5-based `SummarizedExperiment` object to/from disk.
- The `realize` generic function in the `DelayedArray` package for more information about on-disk realization of objects carrying delayed operations.

Examples

```

nrows <- 200; ncols <- 6
counts <- matrix(runif(nrows * ncols, 1, 1e4), nrows)
colData <- DataFrame(Treatment=rep(c("ChIP", "Input"), 3),
                    row.names=LETTERS[1:6])
se0 <- SummarizedExperiment(assays=SimpleList(counts=counts),
                          colData=colData)

se0
dim(se0)
dimnames(se0)
assayNames(se0)
head(assay(se0))
assays(se0) <- endoapply(assays(se0), asinh)
head(assay(se0))

rowData(se0)
colData(se0)

se0[, se0$Treatment == "ChIP"]

## cbind() combines objects with the same features of interest
## but different samples:
se1 <- se0
se2 <- se1[,1:3]
colnames(se2) <- letters[1:ncol(se2)]
cmb1 <- cbind(se1, se2)
dim(cmb1)
dimnames(cmb1)

## rbind() combines objects with the same samples but different
## features of interest:
se1 <- se0
se2 <- se1[1:50,]
rownames(se2) <- letters[1:nrow(se2)]
cmb2 <- rbind(se1, se2)
dim(cmb2)
dimnames(cmb2)

## -----
## ON-DISK REALIZATION
## -----
setRealizationBackend("HDF5Array")
cmb3 <- realize(cmb2)
assay(cmb3, withDimnames=FALSE) # an HDF5Matrix object

```

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